

Serum interleukins and urinary microglobulin in children with idiopathic nephrotic syndrome

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المستويات المصلية للإنترلوكين 1 - بيتا، والإنترلوكين 6، والعامل المُنتَجَر للورم، ومستوى البيتا 2 - مكروغلوبولين في البول، في الأطفال المصابين بمتلازمة كلأئية مجهولة السبب محمد كامل رزق، أحمد النواوي، إلهام عبد الكريم، السيد عامر، دلال الجزائري، أحمد زكي الشافعي

الخلاصة: درس الباحثون 60 طفلاً مصاباً بالمتلازمة الكلأئية المجهولة السبب، مع عشرين آخرين بمائلونهم من حيث العمر والجنس على أنهم شواهد. وقد قسم الأطفال المصابون بالمتلازمة الكلأئية المجهولة السبب إلى ثلاث مجموعات، كلٌّ منها تضم 20 طفلاً، في الأولى من شوهدهم لأول مرة، وفي الثانية من هم في فترة هدأة، وفي الثالثة من هم في فترة نكس. وأجرى الباحثون دراسة متكاملة لعناصر الدم والبول، مع تحليل لمستويات الإنترلوكين 1 - بيتا، والإنترلوكين 6، والعامل المُنتَجَر للورم، وعبارة كميات بيتا 2 - مكروغلوبولين المفرغ في البول. وقد وجد الباحثون أن مستوى الإنترلوكين 1 - بيتا والإنترلوكين 6، مرتفع لدى المجموعات المدروسة، ويبلغ أقصاه لدى الذين شوهدهم لأول مرة والذين نكسوا. أما مستوى العامل المُنتَجَر للورم، والمفرغ من البيتا 2 - مكروغلوبولين في البول فقد كان أكثر ارتفاعاً لدى من شوهدهم لأول مرة والناكسين بشكل واضح. وبناءً على ذلك فإن تراكيز كل من الإنترلوكين 1 - بيتا والإنترلوكين 6 والعامل المُنتَجَر للورم، يمكنها أن تحدّد تحديداً إيجابياً (بنسبة مئة بالمئة) مجموعات الذين شوهدهم لأول مرة ومجموعات النكس، فيما تحدّد هذه التراكيز إضافة إلى المفرغ من البيتا 2 - مكروغلوبولين في البول تحديداً سلبياً المجموعة الشاهدة بنسبة مئة بالمئة.

ABSTRACT We studied 60 children affected with idiopathic nephrotic syndrome (INS) plus 20 age and sex matched controls. The children with INS were divided into 3 groups of 20: first presentation, remission and relapse. A complete blood picture and complete urinalysis were done. Serum interleukin (IL)-1 β , IL-6, tumour necrosis factor (TNF) and quantitative urinary β -2-microglobulin (β -2-m) excretion were estimated. IL-1 β and IL-6 were significantly higher in the study groups, the first presentation and relapse groups having the highest concentrations. Serum TNF concentration and urinary β -2-m excretion were significantly higher in the first presentation and relapse groups. Serum IL-1 β , IL-6 and TNF concentrations were able to select positively (100%) the first presentation and relapse groups, while these plus urinary β -2-m excretion selected negatively (100%) the control group.

Interleukines sériques et β microglobuline urinaire chez des enfants présentant un syndrome néphrotique idiopathique

RÉSUMÉ Nous avons étudié 60 enfants présentant un syndrome néphrotique idiopathique plus 20 témoins appariés selon l'âge et le sexe. Les enfants présentant un syndrome néphrotique idiopathique ont été répartis en trois groupes de 20 : première manifestation, rémission et récurrence. Un hémogramme et un examen des urines complets ont été réalisés. On a estimé l'interleukine 1 β , l'interleukine 6, la cachectine (TNF) et l'excrétion quantitative de β 2-microglobuline urinaire. Les taux sériques d'interleukine 1 β et d'interleukine 6 étaient significativement plus élevés dans les groupes de l'étude, les groupes de première manifestation et de récurrence ayant les concentrations les plus fortes. La concentration sérique de TNF et l'excrétion de β 2-microglobuline urinaire étaient significativement plus élevées dans les groupes de première manifestation et de récurrence. Les concentrations sériques d'interleukine 1 β , d'interleukine 6 et de TNF permettaient de sélectionner positivement (100 %) les groupes de première manifestation et de récurrence alors que ces concentrations plus l'excrétion urinaire de β 2-microglobuline urinaire sélectionnaient négativement (100 %) le groupe témoin.

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Introduction

Nephrotic syndrome is neither a single disease nor even a heterogeneous group of related diseases. Rather, it is a clinical state characterized by heavy proteinuria and hypoalbuminaemia, often associated with oedema, hypercholesterolaemia and generalized hyperlipidaemia [1].

Idiopathic nephrotic syndrome (INS) accounts for 90% of nephrosis in childhood; minimal change nephrotic syndrome is found in approximately 85% of INS, mesangial proliferative glomerulonephritis in 5% and focal glomerular sclerosis in 10%. In the remaining 10% of children with nephrotic syndrome, it is largely mediated by some form of glomerulonephritis, membranous and proliferative being the most common [2].

Idiopathic nephrotic syndrome in childhood occurs at an annual incidence of 2 cases/100 000 population under the age of 18 years [3]; minimal change nephrotic syndrome is most common in children 2–4 years old, but may occur at any age [4].

The basic pathogenic abnormality in nephrosis is proteinuria, which results from an increase in glomerular capillary wall permeability. The mechanism of the increase in permeability is unknown but may be related, at least in part, to loss of negatively charged glycoproteins within the capillary wall [2]. The glomerular lesion, especially in minimal change nephrotic syndrome, may be mediated by circulating permeability factors. They are thought to be T-lymphocyte derived cytokines, which cause podocyte swelling, alterations in charge density, and foot process effacement with consequent increase in vascular permeability [5].

Interleukin (IL)-1 and tumour necrosis factor (TNF) are structurally unrelated cytokines, yet their spectra of biological ef-

fects are so similar that these 2 cytokines are almost interchangeable [6]. It has been suggested that IL-1 has a significant role in the immunopathogenesis of proteinuria [7] and that TNF- α plays a pathogenic role in the induction and/or maintenance of glomerular barrier dysfunction [8]. Interleukin-6 is involved in inflammatory responses and immune reactions in the host. It is produced by a variety of cells, including monocytes and mesangial cells in the kidney [9]. It is not possible at present to distinguish whether IL-6 contributes to renal dysfunction or whether it reflects renal damage [10].

β -2-microglobulin (β -2-m) is a low molecular weight protein found on the surface of all nucleated cells which synthesize it. About 95% of free β -2-m is filtered by the normal glomerulus and a normal kidney is able to reabsorb 99.9% through the proximal tubules. Malfunction of the proximal tubules with a normal glomerular filtration rate is accompanied by decreased tubular reabsorption and increased urinary excretion of β -2-m [11].

The aim of this study was the evaluation of serum concentrations of IL-1 β , IL-6, TNF and urinary β -2-m in children suffering from idiopathic nephrotic syndrome in various situations: before treatment, during remission and on relapse before reinstatement of therapy. An additional aim was the comparison of these levels to those in a matched control group.

Methods

This study was carried out at Alexandria University Children's Hospital from January 1998 to December 1999 (2 years).

Eighty children were included in the study, 60 had idiopathic nephrotic syndrome (INS) and 20 were healthy children who were randomly selected from those

attending the growth and development follow-up clinic who were included as a control group. The sick children were recruited in the paediatric nephrology clinic and selected sequentially until the required number was reached. They were classified into 3 groups (20 children in each): first presentation, diagnosed as nephrotic for the first time (these were selected first owing to their small number); remission, selected as being in remission for ≥ 3 months; and relapse, selected as having had their first relapse recently but before reinstitution of therapy (after the first group was complete, we selected a child in remission and a child with relapse sequentially until the required number was reached). Informed consent was obtained from all parents and the protocol of the study was approved by the Alexandria University ethics committee. All patients with a history of recent (within the previous 6 months) infective and/or inflammatory conditions and all patients with abnormal urinary sediments (abnormal casts or crystalluria) were excluded from the study.

A detailed history of the present condition of all the participants was obtained and a thorough clinical examination was performed.

A morning blood sample of 8 mL was collected; 2 mL of the sample were added to EDTA tubes for a complete blood picture. The remaining 6 mL were collected in test tubes for clinical tests. Sera were separated and collected in Eppendorf tubes. These were kept frozen (-70°C) till the time of the specific tests. All investigations were carried out in the laboratories of the university hospital.

The specific tests performed were: complete blood picture, blood urea, serum creatinine, serum protein and serum cholesterol. Specific immunologic tests included: serum IL-1 β by enzyme immunoassay u-

sing Medgenix kits (Medgenix Diagnostics, Fleurus, Belgium) [intra-assay coefficient of variation (CV) 2.8%, interassay CV 4.5%]; serum IL-6 by enzyme immunoassay using Medgenix kits (intra-assay CV 5.1%, interassay CV 4.6%); TNF- α by enzyme immunoassay using Medgenix kits (intra-assay CV 4.8%, interassay CV 8.7%). All tests were carried out according to the manufacturer's instructions.

An early morning sample of urine was collected, examined and centrifuged. The clear supernatant was preserved for the definitive test. Urine output was estimated by collecting all urine voided from 08.00 till 08.00 the next day. For the control group, mothers were asked to collect the urine and deliver it to the clinic for analysis. β -2-m was estimated in urine quantitatively using an enzyme-linked immunosorbent assay technique using Eurogenetics kits coated microtitre strips (Eurogenetics, Tessenderlo, Belgium) (intra-assay CV 6.4%, inter-assay CV 8.0%).

Data were collected and tabulated using SPSS, version 6, for statistical analysis. Descriptive measures included: count, percentage, minimum, maximum, arithmetic mean and standard deviation. Statistical tests included chi-squared for testing association, 1-way analysis of variance (F -test) for comparing means of more than 2 groups, Tukey β difference test for pair-wise comparison and Kruskal-Wallis analysis of variance (χ^2) for non-parametric or non-normally distributed variables. Pearson correlation was utilized to study association among quantitative variables. The selected level of significance was $P \leq 0.05$.

Results

There were no statistically significant differences between the 4 groups for mean age,

sex distribution and height percentile from 50th centile for age and sex. The weight percentiles were significantly higher in the first presentation and relapse groups than in the control and remission groups from 50th centile for age and sex. Systolic and diastolic blood pressures were significantly higher in the 3 nephrotic groups compared to the control group (Table 1).

Mean serum protein concentration was significantly lower in the 3 study groups, and in the relapse group it was significantly lower than in the other INS groups (Table

2). The serum albumin concentration was significantly lower in the 3 study groups compared to the control group, and also in the first presentation and relapse groups compared to the remission group. Mean serum cholesterol was significantly higher in the 3 INS groups than in the control group. Mean serum creatinine was significantly higher in the first presentation and remission groups compared to the control group, but mean blood urea did not show any statistically significant difference across the 4 groups. The mean leukocyte count was

Table 1 Personal and clinical characteristics of 3 groups of children with nephrosis and a control group

Characteristic	Group				F
	First presentation	Remission	Relapse	Control	
<i>Age (years)</i>					
Range	2.1–11.5	2.3–14.3	2.4–12.6	2.3–10.2	
Mean (SD)	5.7 (3.0)	6.7 (3.4)	5.7 (3.8)	4.0 (2.2)	2.4543
<i>Sex</i>					
Males [No. (%)]	11 (55)	11 (55)	14 (70)	8 (40)	$\chi^2 = 3.636^a$
Females [No. (%)]	9 (45)	9 (45)	6 (30)	12 (60)	
<i>Weight percentile^b</i>					
Range	116.4–135.1	76.5–101.7	81.2–194.5	79.3–101.8	
Mean (SD)	126.5 (18.1) ^{c,d}	94.0 (6.3)	125.7 (31.8) ^{c,d}	92.6 (6.1)	20.255 ^e
<i>Height percentile^a</i>					
Range	84–144.8	86.8–160.0	84–149.7	84–138.3	
Mean (SD)	111.8 (19.0)	118.3 (20.6)	111.1 (23.9)	101.2 (14.7)	2.5238
<i>Systolic blood pressure (mmHg)</i>					
Range	100–150	100–140	80–140	90–110	
Mean (SD)	105.5 (11.8) ^d	103.1 (12.2) ^d	121 (16.8) ^d	92.5 (5.5)	18.425 ^e
<i>Diastolic blood pressure (mmHg)</i>					
Range	60–80	60–85	50–100	60–70	
Mean (SD)	65.5 (7.2) ^d	64.2 (6.8) ^d	73.75 (12.9) ^d	60.5 (2.2)	9.323 ^e

^aKruskal–Wallis test.

^bPresented as % from 50th centile for age and sex.

^cSignificantly different from remission group.

^dSignificantly different from control group.

^eSignificant at $P < 0.05$.

SD = standard deviation.

Table 2 Laboratory results from blood and urine tests for 3 groups of children with nephrosis and a control group

Test	Group				F
	First presentation	Remission	Relapse	Control	
<i>Serum protein (g/dL)</i>					
Range	3.3–7.4	5.4–8.2	2.8–7.2	7.0–8.2	
Mean (SD)	6.0 (0.9) ^a	6.1 (1.2) ^a	4.5 (1.3) ^{a,b,c}	7.8 (0.4)	36.092 ^d
<i>Serum albumin (g/dL)</i>					
Range	1.2–2.0	3.1–4.2	1.4–2.0	3.0–4.5	
Mean (SD)	1.7 (0.2) ^{b,a}	3.7 (0.4) ^a	1.8 (0.2) ^{b,a}	3.9 (0.3)	318.000 ^d
<i>Serum cholesterol (mg/dL)</i>					
Range	120–640	140–540	216–561	145–200	
Mean (SD)	401.9 (137.8) ^a	350.0 (129.0) ^a	402.9 (105.6) ^a	168.0 (15.3)	20.051 ^d
<i>Blood urea (mg/dL)</i>					
Range	17–51	18–51	10–56	20–35	
Mean (SD)	34.2 (10.3)	29.9 (8.6)	32.5 (14.1)	25.9 (4.3)	2.603
<i>Serum creatinine (mg/dL)</i>					
Range	0.2–1.2	0.5–1.1	0.5–1.2	0.3–0.9	
Mean (SD)	0.9 (0.2) ^a	0.8 (0.2)	0.8 (0.3) ^a	0.6 (0.2)	5.669 ^d
<i>Leukocytes (× 1000/mm³)</i>					
Range	5.7–12.3	6.9–16.2	5.4–20.7	6.2–9.4	
Mean (SD)	8.9 (1.6)	10.0 (2.6) ^a	11.4 (3.9) ^{a,c}	7.8 (1.2)	7.589 ^d
<i>Haemoglobin (g/dL)</i>					
Range	8.0–13.7	9.6–14.0	7.9–15.6	8.3–13.8	
Mean (SD)	10.4 (1.6)	11.8 (1.3)	12.2 (2.2) ^c	11.0 (1.6)	4.359 ^d
<i>Platelets (× 1000/mm³)</i>					
Range	180–693	211–630	280–544	238–430	
Mean (SD)	349.8 (103.2)	369.7 (105.1)	420.7 (71.4) ^a	314.8 (66.3)	5.019 ^d
<i>Leukocytes in urine/HPF</i>					
Range	1–100	1–9	2–55	1–2	
Mean (SD)	17.4 (22.1) ^a	3.45 (2.0) ^c	12.3 (13.1) ^a	1.36 (0.5)	6.856 ^d
<i>Urinary volume (mL/kg per day)</i>					
Range	9.1–125.0	24.0–150.0	25.0–38.5	50.0–68.2	
Mean (SD)	35.8 (27.1) ^{a,b}	60.5 (35.8)	31.0 (3.3) ^{a,b}	60.4 (4.4)	9.625 ^d

^aSignificantly different from control group.^bSignificantly different from remission group.^cSignificantly different from first presentation group.^dSignificant at P < 0.05.

SD = standard deviation.

HPF = high power field.

significantly higher in the remission and relapse groups compared to the controls, and in the relapse group it was significantly higher than in the first presentation group. Mean platelet count was significantly higher in the relapse group compared to controls and mean haemoglobin concentration was significantly higher in the relapse group compared to the first presentation group.

Mean urinary leukocyte count was significantly higher in the first presentation and relapse groups compared to the control group, while in the first presentation group the count was significantly higher than in the remission group. Mean 24-hour urinary output was significantly lower in the first presentation and relapse groups compared to the other 2 groups.

Mean serum IL-1 β concentration was significantly higher in the first presentation and relapse groups compared to the remission and control groups, while in the remission group the level was significantly higher than in the controls (Table 3). Mean serum IL-6 concentration was significantly higher in the 3 groups with nephrosis compared to the controls, and in the first presentation and relapse groups the level was significantly higher than the remission group. Mean serum TNF concentration in the first presentation and relapse groups was significantly higher than that of the remission and control groups, and in the first presentation group it was significantly higher than in the relapse group. Mean urinary β -2-m concentration in the first presentation and relapse groups was

Table 3 Serum concentrations of interleukin (IL)-1 β , IL-6, tumour necrosis factor (TNF) and urinary β -2-microglobulin (β -2-m) in children with nephrosis and a control group

Variable	Group				Kruskal-Wallis test, χ^2
	First presentation	Remission	Relapse	Control	
<i>IL-1β (pg/mL)</i>					
Mean (SD)	1208.2 (1977.5)	73.4 (91.6)	657.3 (790.5)	6.0 (2.7)	59.74 ^a
Negative [No. (%)]	0 (-)	8 (40)	0 (-)	20 (100)	
Positive [No. (%)]	20 (100)	12 (60)	20 (100)	0 (-)	58.90 ^a
<i>IL-6 (pg/mL)</i>					
Mean (SD)	448.5 (268.3)	32.0 (40.6)	171.3 (182.1)	4.1 (2.2)	
Negative [No. (%)]	0 (-)	7 (35)	0 (-)	20 (100)	65.682 ^a
Positive [No. (%)]	20 (100)	13 (65)	20 (100)	0 (-)	59.65 ^a
<i>TNF (pg/mL)</i>					
Mean (SD)	56.6 (9.3)	29.3 (58.8)	43.9 (24.6)	5.6 (2.8)	51.424 ^a
Negative [No. (%)]	0 (-)	12 (60)	0 (-)	20 (100)	60.00 ^a
Positive [No. (%)]	20 (100)	8 (40)	20 (100)	0 (-)	
<i>Urinary β-2-m (μg/mL)</i>					
Mean (SD)	6.0 (5.2)	2.5 (4.9)	4.9 (7.6)	1.4 (0.9)	21.86 ^a
Negative [No. (%)]	13 (65)	17 (85)	9 (45)	20 (100)	17.76 ^a
Positive [No. (%)]	7 (35)	3 (15)	11 (55)	0 (-)	

^aSignificant at $P < 0.05$.

Negative = \leq normal.

Positive = $>$ normal.

SD = standard deviation.

significantly higher than in the remission and control groups.

Serum concentrations of IL-1 β , IL-6 and TNF were able to select positively the first presentation and relapse groups (100%), while these 3 cytokines along with urinary β -2-m excretion could select negatively the controls (100%).

Discussion

Increased level of IL-1 β encountered in a study is indicative of its significant role in the immunopathogenesis of proteinuria [7]. The effect of increased IL-1 β has been explained on the basis of its local and systemic effects. The main vascular effects of IL-1 β are increased platelet adherence, increased capillary permeability, increased prostaglandin synthesis and hypertension [12]. Moreover, IL-1 β with TNF may stimulate increased synthesis of eicosanoids [2]. These could pass through the endothelial cells, affecting the negatively charged podocytes, which usually prevent the passage of albumin, leading to their neutralization and hence excessive albuminuria.

Although the mean blood pressure measurements among the INS groups in our study were significantly higher than that of the control group, they were within the normal range for age and sex in the majority of children [2]. It has been postulated that increased mesangial cellularity [13] could compromise renal afferent and/or efferent circulation in the glomeruli. Abrass reported that 19% of children remain hypertensive after induction of remission in nephrotic syndrome [14].

An important effect of increased IL-1 β combined with IL-6 and TNF is the increased leukocyte count from 2 sources: bone marrow and leucocytes attached loosely to endothelial cells [15]. This high

leukocyte count could possibly indicate that the infiltrating monocytes-macrophages are the major source of inflammatory cytokines, especially IL-1 β , IL-6 and TNF, rather than the resident glomerular cells. IL-1 β stimulates hepatic protein synthesis due to hypoalbuminaemia and inhibits lipoprotein lipase [12], which will lead to hypercholesterolaemia. In this study, the main cause of hypercholesterolaemia in the remission group was inhibition of lipoprotein lipase since the serum protein was within the normal range for age and sex [2]. Although IL-1 β increased hepatic protein synthesis, its vascular leak effects surpass it, with a net hypoteinaemia.

The expression of TNF is amplified by IL-1 β and is suppressed by immunosuppressive drugs such as corticosteroids [16]. In renal disease, TNF expression has been found in both resident cells and infiltrating monocytes-macrophages [17]. It acts in a paracrine way to recruit monocytes and macrophages to the glomerular region, besides acting with other mediators to increase vascular permeability [18]. This would cause alteration of the barrier function of the capillary wall [17] leading to proteinuria. In our study, TNF did not behave similar to IL-1 β in the remission group, where TNF levels decreased to statistically non-significant levels compared to controls while IL-1 β levels remained significantly high. This is because TNF is downregulated by corticosteroids [17] and also by the shift of CD₄-T cell differentiation to TH₂ instead of TH₁ cells with a consequent reduction of TH₁ cytokine production, especially TNF [19]. Moreover, the combined effect of IL-1 β and TNF will lead to a more effective local capillary leakage and cell death than with either cytokine alone [20]. Therefore, to obtain remission there should be a disengagement of IL-1 β and TNF. This emulates

IL-1 β and IL-6 in its systemic effects, especially bone marrow stimulation and release of neutrophils [15].

While IL-1 β and TNF are stimulating mesangial cells to release IL-6 [17,20], other sources are activated: TH₂ cells, antigen presenting cells and other somatic cells, especially endothelial cells [12]. Concentration of IL-6 was highest in the first presentation group and lowest in the remission group in the INS patients in this study. IL-6 is activated by the high IL-1 β and TNF concentrations in the first presentation group, thus IL-6 seems to be able to downregulate TNF, possibly by helping in differentiating CD₄-T cells into TH₂ cells. Consequently, remission will occur and IL-6 will no longer be needed in the remission group and concentration will ultimately decline. This view is supported by the findings that long-term remission of INS is related to the increased TH₂ cytokine production (especially IL-6) and the downregulation of TH₁ cells [19]. Proteinuria disappears in remission since it has been shown that TNF alone is the single most important cytokine in pathogenesis in INS [21]. Moreover, the use of corticosteroids in the remission group suppressed IL-6 [22]. In the relapse group in this study, the proteinuria caused by increased TNF will need a re-increase of IL-6 expression to suppress TNF expression and its proteinuric effect.

In this study, IL-6 combined with other cytokines acted on the bone marrow to increase leukocyte count and haematopoiesis [12] as well as megakaryocytes [23]. The effect is latent and therefore the platelets increased in the relapse group while the leukocyte count depended more on the effects of other cytokines such as IL-1 β and TNF, which can be seen from the results.

The lowest haemoglobin concentration we found was in the first presentation group, possibly as a result of low level of

erythropoietin in plasma which can result from increased urinary loss in nephrotic syndrome [14]. This may have outweighed the stimulatory effects of erythropoietin production in bone marrow by the high concentrations of IL-1 β and IL-6 in the first presentation group, but the reverse occurred in the relapse group.

In this study, the highest increase of urinary excretion of β -2-m was in the first presentation group because of the combined effect of glomerular and tubular dysfunction. It was shown that increased urinary β -2-m excretion is present in massive glomerular proteinuria and in tubular dysfunction, especially the proximal [24], with a positive correlation between them independent of the primary renal disease [25].

To sum up, significantly high concentrations of IL-1 β , IL-6 and TNF were uniformly found in the first presentation and relapse INS groups. They were able to positively predict them (100%) and, along with β -2-m, to negatively predict the controls (100%) with fair accuracy. The lower serum values for TNF and urinary β -2-m excretion are important factors in defining and predicting remission. IL-1 β and TNF usually work in harmony while IL-6 tries to suppress TNF and its harmful effects. It seems that the 3 months remission period used in this study is not a sufficient duration for the return of cytokines and urinary β -2-m to the basal control values. However, the clinical studies on INS in children are quite limited because of its low global incidence (2/100 000 population) [3].

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References

1. Nash MA et al. The nephrotic syndrome. In: Edelman CM ed. *Pediatric kidney disease*, 2nd ed. Boston, Little Brown and Company, 1992:1247–66.
2. Bergstein JM. Conditions particularly associated with proteinuria. In: Behrman RE, Kliegman RM, Jenson HB, eds. *Nelson textbook of pediatrics*, 16th ed. Philadelphia, WB Saunders, 2000:1590–6.
3. Salgado JR et al. Nephrosis in childhood. *Nephron*, 1995, 71(4): 373–85.
4. Ordonez N, Rosai J. Urinary tract. In: Rosai J, ed. *Ackerman's surgical pathology*, 8th ed. St Louis, CV Mosby, 1996: 1059–163.
5. Gibson IW, More IA. Glomerular pathology: recent advances. *Journal of pathology*, 1998, 184(4):123–9.
6. Oppenheim JJ, Ruscetti FW, Faltynek C. Cytokines: complement & kinin. In: Stites DP, Stobo JD, Wells VJ, eds. *Basic and clinical immunology*, 8th ed. Norwalk, Connecticut, Appleton & Lange, 1994: 105–23.
7. Saxena S, Mittal A, Andal A. Pattern of interleukins in minimal-change nephrotic syndrome of childhood. *Nephron*, 1993, 65(1):56–61.
8. Suranyi MG et al. Elevated levels of tumour necrosis factor- α in the nephrotic syndrome in humans. *American journal of kidney disease*, 1993, 21(3):251–9.
9. Ohta V et al. Detection and clinical usefulness of urinary interleukin-6 in the diseases of the kidney and the urinary tract. *Clinical nephrology*, 1992, 38(4):185–9.
10. Akutsu Y. [Study of correlation between urinary IL-6 level and mesangial lesion in childhood onset Ig A nephropathy]. *Hokkaido igaku zasshi*, 1994, 69(4): 686–96 (in Japanese).
11. Cohen JJ, Harrington JT, Madias NE. Dialysis related amyloidosis. *Kidney international*, 1992, 41:1416–29.
12. Ganong WF. Circulating body fluids. In: Ganong WF, ed. *Review of medical physiology*, 19th ed. Stamford, Connecticut, Appelton & Lange, 1999, 493–521.
13. Tesch GH et al. Role of interleukin-1 in mesangial cell proliferation and matrix deposition in experimental mesangioproliferative nephritis. *American journal of pathology*, 1997, 151(1):141–50.
14. Abrass CK. Clinical spectrum and complications of the nephrotic syndrome. *Journal of investigative medicine*, 1997, 45(4): 143–53.
15. Janeway CA et al. T-cell mediated immunity. In: Janeway CA et al., eds. *Immunobiology: the immune system in health and disease*, 4th ed. New York, Elsevier Science, 1999:363–415.
16. Baud L, Ardaillou R. Tumour necrosis factor in renal injury. *Mineral and electrolyte metabolism*, 1995, 21:336–41.
17. Abott F et al. Interleukin-1 β stimulates human mesangial cells to synthesize and release interleukins-6 and -8. *Kidney international*, 1991, 40(4):597–605.
18. Gruss HJ. Molecular, structural and biological characteristics of the tumour necrosis factor ligand superfamily. *International journal of clinical & laboratory research*, 1996, 26(3):143–59.
19. Daniel V et al. T-lymphocyte populations, cytokines and other growth factors in serum and urine of children with idiopathic nephrotic syndrome. *Clinical nephrology*, 1997, 47(5):289–97.
20. Dinarello CA. Reduction of inflammation by decreasing production of interleukin-1 or by specific receptor antagonism. *Inter-*

- national journal of tissue reactions*, 1992, 14(2):65–75.
21. Kluth DC, Rees AJ. New approaches to modify glomerular inflammation. *Journal of nephrology*, 1999, 12(2):66–75.
22. Fukatsu A et al. Expression of interleukin-6 and major histocompatibility complex molecules in tubular epithelial cells of diseased human kidneys. *Laboratory investigation*, 1993, 69(1):58–67.
23. Hirano T. Interleukin-6 and its receptor: ten years later. *International reviews of immunology*, 1998, 16(3–4):249–84.
24. Fede C et al. N-acetyl- β -D-glucosaminidase and beta-2-microglobulin: prognostic markers in idiopathic nephrotic syndrome. *Journal of nephrology*, 1999, 12(1):51–5.
25. Caleskan S et al. Urinary N-acetyl- β -D-glucosaminidase and β -2-microglobulin excretion in primary nephrotic syndrome in children. *Nephron*, 1994, 74(2):401–4.

Note from the Editor

We would like to inform our readers that the next special issue of the EMHJ will be on Medical Bioethics. It is scheduled to be Vol. 12 No. 5, 2006.